# Effect of hydropriming on morphological and physiological performance of aged groundnut (*Arachis hypogaea* L.) seeds

Ali Sepehri<sup>1\*</sup> and Hossein Reza Rouhi<sup>2</sup>

1, 2. Associate Professor and Ph.D. Candidate, Department of Agronomy and Plant Breeding, Faculty of Agriculture, Bu-Ali Sina

University, Hamedan, Iran

(Received: Feb. 8, 2016 - Accepted: Nov. 12, 2016)

#### ABSTRACT

Low viability and poor vigour due to seed deterioration are major problems for successful agricultural production. In this experiment, the ability of hydropriming to ameliorate seed deterioration damage was studied in groundnut. Groundnut seeds were subjected to accelerated ageing for 96 hours at 40 °C and then hydroprimed at 25 °C for 6, 12 and 18 hours. Regardless of duration, hydropriming significantly improved final germination percentage, germination rate, seedling length, vigour index, antioxidant enzyme activities (catalase, superoxide dismutase and ascorbate peroxidase), soluble sugars and proteins of aged seeds. Mean germination time, electrolyte leakage and malondialdehyde content of primed seeds decreased compared to non-primed seeds. Hydropriming for 6, 12 and 18 hours increased final germination to 15.54, 31.56 and 89.7 %, germination rate by 9.42, 66.6 and 95.6 % and vigour index by 29.11, 82.2 and 204.6 % compared to non-primed seed, respectively. Thus, hydropriming for 18 hours is the most suitable priming period to recover loss of seed quality and to improve germination characteristics of aged groundnut seeds.

Keywords: Antioxidant enzyme activities, seed germination, seedling length, soluble sugars.

# اثر هیدرو پر ایمینگ بر کار آیی مورفولوژی و فیزیولوژی بذور زوال یافته بادام زمینی (Arachis hypogaea L.) علی سپهری<sup>ا\*</sup> و حسین رضا روحی<sup>۲</sup> ۱ و ۲. دانشیار و دانشجوی دکتری، گروه زراعت و اصلاح نباتات، دانشکده کشاورزی، دانشگاه بوعلی سینا، همدان

(تاریخ دریافت: ۱۳۹٤/۱۱/۱۹ – تاریخ پذیرش: ۱۳۹۵/۸/۲۲)

#### چکیدہ

زوال بذر یکی از مشکلات اصلی در تولید موفق محصولات کشاورزی به علت کاهش قوهنامیه و ضعف بنیه بذر است. در این آزمایش توانایی هیدروپرایمینگ بذر در تخفیف خسارت ناشی از زوال بذر بادام زمینی بررسی شد. بذرهای بادام زمینی یه روش پیری زودرس بهمدت ۹۲ ساعت در دمای ٤٠ درجه سانتی گراد زوال یافتند. سپس بذور زوال یافته در دمای ٢٥ درجه سانتی گراد بهمدت ٦، ١٢ و ١٨ ساعت هیدروپرایم شدند. صرفنظر از مدتزمان، هیدروپرایمینگ به طور معنی داری منجربه بهبود درصدجوانهزنی، سرعت جوانهزنی، طول گیاهچه، شاخص بنیه، فعالیت آنزیمهای آنتی اکسیدانی (کاتالاز، سوپراکسید دیسموتاز و آسکوربات پراکسیداز)، قندها و پروتئینهای محلول بذور زوال یافته گردید. متوسط زمان جوانهزنی، هدایت الکتریکی و محتوای مالون دی آلدهید بذور پرایم شده در مقایسه با بذور پرایم نشده کاهش یافت. هیدروپرایمینگ به مدت ٦، ١٢ و ١٨ ساعت درصد جوانهزنی نهایی را ۱۵/۵۲، ۱۸/۷ مول ساعت در مقایسه با بذور زوال یافته گردید. متوسط زمان جوانهزنی، هدایت الکتریکی و محتوای مالون دی آلدهید بذور پرایم شده در مقایسه با بذور پرایم نشده کاهش یافت. هیدروپرایمینگ به مدت ٦، ١٢ و ١٨ ساعت درصد جوانهزنی نهایی را ۱۵/۵۱، ۱۰/۳، ۷/۷ مرعت جوانهزنی را ۱۹/۵۲، ۱/۲۵، ۲۰/۱۹ و شاخص بنیه را به ترتیب ۱۹/۱۲، ۲/۸۲، ۲/۱۸ ۲/۵۶ در مقایسه با بذور پرایم نشده افزایش داد. بنابراین، هیدروپرایمینگ به مدت ۱۸ ساعت به عنوان مناسب ترین زمان پرایمینگ جهت بازیابی کیفیت از دست رفته بذور و بهبود خصوصیات جوانهزنی بذور زوال یافته بادام زمینی می باشد.

واژههای کلیدی: آنزیم های آنتی اکسیدان، جوانهزنی بذر، طول گیاهچه، قندهای محلول.

\* Corresponding author E-mail: sepehri110@yahoo.com; a\_ sepehri@basu.ac.ir

#### Introduction

Groundnut (Arachis hypogaea L.) is one of the most important oilseed crops in the Fabaceae; however, oil in the seeds are easily oxidized during storage condition, resulting in decreased seed viability and vigour. Low vigour seed is the cause of poor crop establishment and thus a major limitation of grain vield (Jisha et al., 2013). Seed priming has been developed as an essential technique to enhance seed performance against oxidative stress (Ashraf & Foolad, 2005; Jisha et al., 2013; Yan, 2015). It has been reported that hydropriming is the simplest, safest and the most economical technique for increasing seed performance, seedling establishment and crop production under unfavorable conditions (Ashraf & Foolad, 2005; Jisha et al., 2013). The efficacy of hydropriming depends on factors such as plant species, priming period and seed vigour (Ahmad & Lee, 2011). Ghasemi-Golezani et al. (2013) showed that hydropriming increased plant height, number of pods and number of seeds per pod, 1000-seed mass, biological yield, grain yield and harvest index of lentil (Lens culinaris) seeds when compared to the control group. Ahmad et al. (2014) emphasized that hydropriming is the cheapest approach to hydrating seeds and minimizes the use of chemical agents. Further, morphological, physiological biochemical parameters and are enhanced in many crops after seed hydropriming (Wattanakulpakin et al., 2012; Yan, 2015; Kamithi et al., 2016; Lopez et al., 2016). Also, these studies have shown that priming is accompanied with the improvement of antioxidant enzyme systems and the accumulation of soluble sugars and soluble proteins. However, physiological morphological and changes induced by seed priming have been rarely analyzed in seedlings. The

objective of this study was to evaluate the effects of hydropriming on the morphological and physiological performance of aged groundnut seeds.

# Materials and Methods Site description and plant material

The current study was carried out at the Department of Agronomy and Plant Breeding, Faculty of Agriculture, Bu-Ali Sina University, Iran. Groundnut seeds cv. NC2 were attained from Astaneh-Ashrafieh (Guilan Province).

#### Seed ageing treatment

Seeds were aged with accelerated ageing at  $40\pm1$  °C for 96 hours (Delouche & Baskin, 1973).

#### Seed priming treatment

Groundnut seeds were soaked in distilled water at 25 °C for 6, 12 and 18 hours under dark conditions. The hydroprimed seeds were surface-dried and dried back to their original moisture content at room temperature ( $25\pm2$  °C for 24 hours), calculated by changes in primed seed mass.

#### **Germination tests**

Four replicates of 100 seeds were tested for germination between double layers of papers moistened with 15 ml of distilled water in 15 cm Petri dishes. Seeds were incubated in dark conditions at  $25\pm1$  °C in 12 days. Germination was documented to have occurred when the radicles were 2mm long.

#### **Morphological measurements**

Germination percentage was recorded every day during the incubation period. Mean germination time (MGT) was calculated using the formula of Ellis & Roberts (1981):

$$MGT = \sum Dn/n$$

Where n is the number of seeds that germinated on day D, and D is the

number of days counted from the									
beginning of germination. Germination									
rate (GR) was calculated by reversing									
MGT (Rahnama-Ghahfarokhi &									
Tavakkol-Afshari, 2007).									
Vigour Index =									
seedling length $\times$ germination percentage									
100									

100

#### **Physiological measurements**

Electrolyte leakage was measured according to method of Hampton & TeKrony (1995). After 10 days, seedlings were used to determine malondialdehyde (Cavalcanti et al., 2004), SOD activity (Giannopolitis & Ries, 1977), CAT activity (Cakmak & Horst, 1991), APX activity (Nakano & Asada, 1981). soluble proteins (Bradford, 1976) and soluble sugars (Irigoyen et al., 1992).

### Statistics

The experiment was a completely randomized with design four replications. Data for germination and abnormal germination percentage were subjected to arcsine transformation before analysis of variance was done with SAS software (version 9.1). Mean comparison was performed with an LSD test if the F - test was significant at 5 and 1% probability levels (P < 0.05and P < 0.01). Graphical presentation of data was done with Microsoft Excel program.

## **Results and Discussion**

The initial germination percentage, germination rate and seedling length of groundnut seeds were 89.33 %, 0.286 and 9.18 cm, respectively. After accelerated ageing, the final germination percentage, germination rate and seedling length had diminished 49.33%, 0.138 and 4.8 to cm. respectively (data not shown). All measured parameters were affected by hydropriming treatments (Table 1). Hydropriming increased final germination percentage (Figure 1), germination rate (Figure 3), seedling length (Figure 4), vigour index (Figure 5), total soluble proteins (Figure 7), soluble sugars (Figure 8) and enzyme activities (Figures 10, 11 and 12) but germination time, electrical mean malondialdehyde conductivity and content did not decline (Figures 2, 6 and 9). Final germination percentage and germination rate were significantly increased by increasing priming period 1 and 3). Hydropriming (Figures enhanced the germination percentage and germination rate of pinto bean (Phaseolus vulgaris) seeds (Ghasemi-Golezani et al., 2010) and the germination rate of rice (Oryza sativa) seeds (Matsushima & Sakagami, 2012). Lopez et al. (2016) found that hydropriming increased germination rate and synchrony and reduced the lag time for start of germination of Dodonaea viscosa seeds. thereby successfully invigorating artificially aged seeds. Sallam (1999) demonstrated that water-soaked seeds of Vicia faba exhibited significantly higher germination percentage and rate than those from nontreated seed. They also reported these effects that were correlated with increased amounts of total soluble sugars, lactose and maltose.

By increasing the period of hydropriming, mean germination time of groundnut significantly decreased. The lowest MGT value was observed after hydropriming for 18 hours and the highest was detected in non-primed seeds (Figure 2). The reduction of MGT via seed priming agrees with the results obtained by Yan (2016), who reported that hydropriming reduce MGT and enhanced seedling growth in two napa cabbage cultivars. Sung and Chiu (1995) reported that hydropriming decreased MGT without changing the amount of water imbibed by watermelon (*Citrullus lanatus*) seeds. The extended priming period positively affected length of seedlings from aged seeds (Figure 4). Seedling length increased significantly with an increase in the period of priming from 6 to 18 hours. Ghasemi-Golezani *et al.* (2008) found that hydropriming treatment increased the germination rate and seedling dry weight of lentil (*Lens culinaris*) seeds.

Table 1. Analysis of variance for hydropriming treatment effects on germination traits of aged groundnut seeds (cv. NC2)

Source of	đf	Mean Squares											
variation	df	FGP	MGT	GR	SL	VI	EL	SS	SP	MDA	CAT	SOD	APX
Treatment	3	428.885**	8.838**	0.012**	13.793**	13.755**	26.077**	87.44**	12.298**	305.372**	$0.004^{**}$	32.272**	0.010**
Error	8	8.886	0.031	0.00005	0.096	0.113	0.472	6.789	0.101	1.191	0.00003	0.457	0.00004
C.V. (%)	-	4.79	3.60	3.24	4.84	7.92	2.83	10.24	3.83	3.12	2.23	2.76	1.80
ma ** * m	no ** * non significant and significant at 1 and 5 nereant of muchability, respectively,												

ns, \*\*, \* non-significant and significant at 1 and 5 percent of probability, respectively.

FGP: Final Germination Percentage, MGT: Mean Germination Time, GR: Germination Rate, SL: Seedling Length, VI: Vigour Index, EL: Electrolyte leakage, SS: Soluble Sugars, SP: Soluble Proteins, MDA: Malondialdehyde content, CAT: Catalase, SOD: Superoxide dismutase, APX: Ascorbate peroxidase.

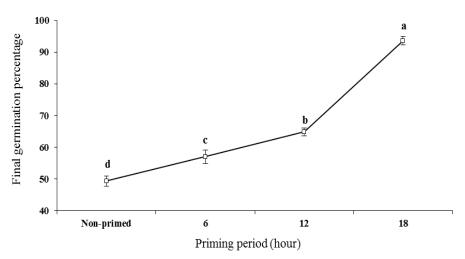


Figure 1. Final germination percentage of aged groundnut seeds in different period of hydropriming

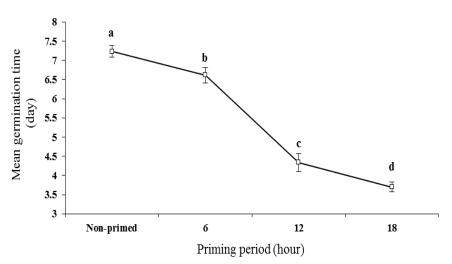


Figure 2. Mean germination time of aged groundnut seeds in different period of hydropriming.

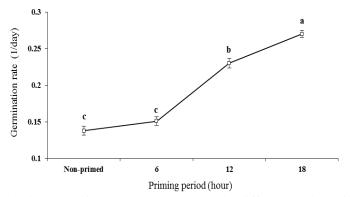


Figure 3. Germination rate of aged groundnut seeds in different period of hydropriming.

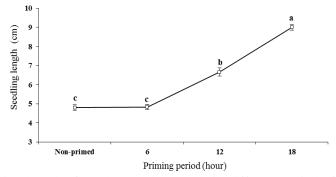


Figure 4. Seedling length of aged groundnut seeds in different period of hydropriming.

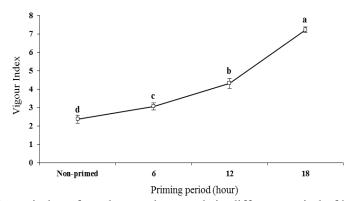


Figure 5. Vigour index of aged groundnut seeds in different period of hydropriming.

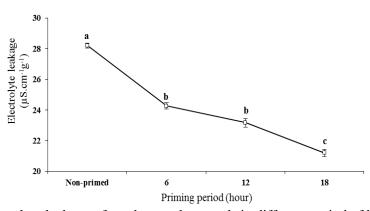


Figure 6. Electrolyte leakage of aged groundnut seeds in different period of hydropriming.

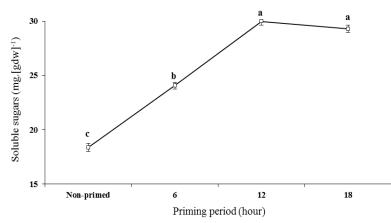


Figure 7. Soluble sugars of aged groundnut seeds in different period of hydropriming

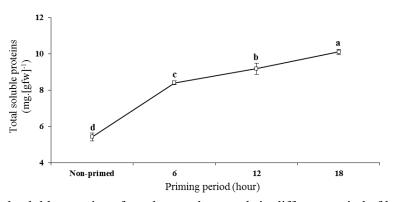


Figure 8. Total soluble proteins of aged groundnut seeds in different period of hydropriming

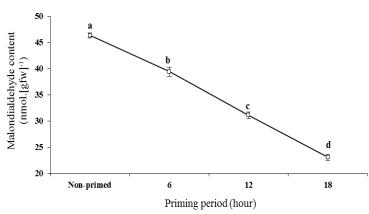


Figure 9. Malondialdehyde content of aged groundnut seeds in different period of hydropriming

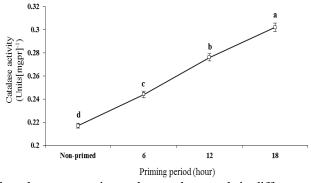


Figure 10. Activity of catalase enzyme in aged groundnut seeds in different period of hydropriming

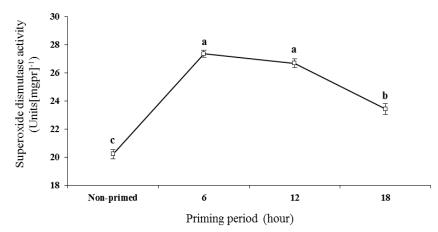


Figure 11. Activity of superoxide dismutase enzyme in aged groundnut seeds in different period of hydropriming

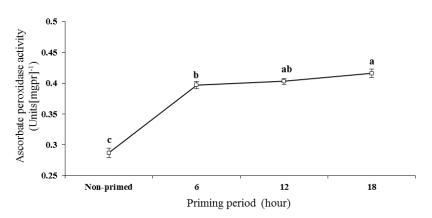


Figure 12. Activity of ascorbate peroxidase enzyme in aged groundnut seeds in different period of hydropriming

In seeds of some plant species, trypsin-like proteolytic enzymes that are produced during seed development are important during germination (Ashraf & Foolad, 2005; Matsushima & Sakagami, 2012; Yan, 2015). The activity of such enzymes, however, is often prevented by trypsin inhibitors, which may be present in the seed and play regulatory roles in protein mobilization during germination (Ashraf & Foolad, 2005). Priming, however, may reduce the inhibitory activities of such enzymes and promote germination as well as cell elongation.

Submersing seeds in water for 18 hours caused a significant increase in the vigour index compared to other priming periods and non-primed (Figure 5). Sharma *et al.* (2014) reported that

hydropriming improved the germination traits especially the vigour index of okra seeds. Priming usually induces faster and more uniform seed germination and seedling vigour index (Nascimento, 2012). Matsushima & Sakagami (2012) demonstrated that hydropriming for 12 h enhanced emergence and seedling vigour of rice seeds under different soil moisture conditions.

Electrolyte leakage decreased after all priming periods (Figure 6). Chiu et al. (1995)indicated that improved membrane repair in primed seeds might cause the better performance of primed seeds because of lower leakge of electrolytes from the cells. Fujikura et al. (1993) also reported the positive effects hydropriming electrical of on conductivity cauliflower of aged

(Brassica oleracea) seeds. Yan (2015) noted that seed ageing induces the accumulation of reactive oxygen species (ROS), which degrade or damage nucleic acids and hydropriming can reduce the negative effects by promoting expression of repaire genes. Soluble sugars and proteins in seedlings from primed seeds were higher than those of the non-primed, while MDA content of seedlings was lower than that of seedlings from nonprimed seeds (Figures 7, 8 and 9). At the cellular level, seed deterioration is with reduced correlated energy metabolism (Kibinza et al., 2006). During priming, carbohydrate reserves in the transformed into seed are simple carbohydrates by hydrolysis of starch. These components are the source of plant energy and are involved in building units (Langens-Grrits et al., 2003; Matsushima & Sakagami, 2012). Thus, the effect of hydropriming improving on seed germination is closely related to the solubilization of b-subunit of 11-S globulin storage protein (Capron et al., 2000). Similarly, in pigeonpea (Cajanus cajan), hydropriming was very effective in the mobilization of compounds such as proteins, free amino acids, and soluble sugars from storage organs to growing embryonic tissues (Jyotsna & Srivastava, 1998).

It seems that free radical production is the leading candidate that causes increasing seed deterioration. Free radical production, primarily triggered by oxygen, has been related to the peroxidation of lipids and degradation of soluble proteins found in cells. This causes a host of undesirable events, including decreased lipid content, reduced respiratory competence, and evaluation of volatile increased compounds such as aldehyde (Wilson & McDonald, 1986). The slow repair of damage membranes during imbibition may cause low germination percentage and seedling vigour (Abdulrahmani et al., 2007). Lowered MDA and EC of primed seed were in accordance with previous studies (Abdulrahmani et al., 2007; Yan, 2015). Reduction in MDA content has been detected by El-Araby & Hegazi (2004) in hydroprimed seeds of tomato. Kamithi et al. (2016) found several processes, including activation synthesis of a number and of antioxidant enzymes and nucleic acids, repair and build up, ATP synthesis, and the cytoplasmic membrane repair, begin during priming. Thus, during priming the content of MDA began to decrease. CAT activity in seeds increased significantly after 6, 12 and 18 hours of priming (Figure 10). The maximum activity of SOD in groundnut seeds was observed after 6 hours of priming (Figure 11). The activity of APX was induced by seed priming, but there was no significant difference between 12 and 18 hours priming (Figure 12).

The deterioration of the aged seed is a natural phenomenon, and seeds tend to lose quality and viability even under optimal storage conditions. There is a correlation between deterioration of seeds during storage and enzymes degradation and inactivation (Bailly, 2004). It is evident that priming can increase antioxidative enzymes such as CAT, APX and SOD in seedlings (Chiu et al., 1995; Sepehri & Rouhi, 2016). A correlation between enhanced activities of antioxidative enzymes and decreased lipid peroxidation has been reported in primed seeds of purple coneflower, bitter gourd and napa cabbage (Hsu et al., 2003; Yan, 2015). Kibinza et al. (2011) noted that CAT is a key enzyme for repairing seeds during priming, and Yan (2015) suggested that CAT activity was correlated with the improvement in germination of primed napa cabbage seeds. The present study, demonstrated hydropriming improved that morphological traits as well as CAT, APX and SOD activities in seedlings,

resulting in a higher final germination and vigour index in aged groundnut seeds. Seed with poor germination parameters showed reduced antioxidant enzyme activity and increased electrolyte leakage associated with cell damage and necrosis, growth of pathogenic fungus, and necrotization of whole seed (Ashraf & Foolad, 2005; Sepehri & Rouhi, 2016). Results from various studies indicate that hydropriming is a useful method for improving the quality of aged seeds, if seed deterioration has not gone too far (Kibinza et al., 2011; Yan, 2015; Kamithi et al., 2016; Yan, 2016). Many investigations have suggested that priming is a practical treatment for increasing germination parameters (Ahmad & Lee, 2011; Ghasemi-Golezani et al., 2013; Ahmad et al., 2014; Yan, 2015). Kaya et al. (2006) increased time found that of hydropriming resulted in higher cumulative germination in sunflower. Also, Caseiro et al. (2004) reported that

hydropriming was the most effective technique for enhancing germination traits of onion seed, especially when seeds were hydrated for 96 h compared to 48 h. According to Sacala & Demczuk (2016), higher time of hydropriming enables seed cells to respond to very low levels of a stimulus in a more rapid and robust manner comparing to non-primed cells.

The results of the current study showed that hydropriming accelerates the germination of groundnut with higher antioxidant enzyme activities, soluble proteins and sugars in seedlings of primed seeds than in those of nonprimed seeds. Priming period had a significant effect on the morphological and physiological traits of groundnut The best results seedlings. were obtained from 18 hours of hydropriming. Therefore, hydropriming for 18 hours can be successfully applied to enhanced seed germination and seedling vigour of aged groundnut seeds.

# REFERENCES

- 1. Abdulrahmani, B., Ghasemi-Golezani, K., Valizadeh, M. & Feizi-Asl, V. (2007). Seed priming and seedling establishment of barley (*Hordeum vulgare* L.). *Journal of Food Agriculture and and Environment*, 5, 179-184.
- 2. Ahmad, G. & Lee, H. C. (2011). Response of sesame (*Sesamum indicum*) cultivars to hydropriming of seeds. *Australian Journal of Basic Applied Science*, 1, 638-642.
- 3. Ahmad, K. U., Rahman, M. M. & Ali, M. R. (2014). Effect of hydropriming method on maize (*Zea mays*) seedling emergence. *Bangladesh Journal of Agriculture Research*, 39(1), 143-150.
- 4. Ashraf, M. & Foolad, M. R. (2005). Pre-sowing seed treatment a shotgun approach to improve germination, plant growth, and crop yield under saline and non-saline conditions. *Advances in Agronomy*, 88, 223-271.
- 5. Bailly, C. (2004). Active oxygen species and antioxidants in seed biology. *Seed Science Research*, 14, 93-107.
- 6. Bradford, M. M. (1976). A dye binding assay for protein. *Analytical Biochemistry*, 72, 248-254.
- 7. Capron, I., Corbineau, F., Dacher, F., Job, C., Come, D. & Job, D. (2000). Sugarbeet seed priming: Effects of priming conditions on germination, solubilization of 11-S globulin and accumulation of LEA proteins. *Seed Science Research*, 10, 243-254.
- 8. Caseiro, R., Bennett, M. A. & Marcos-Filho, J. (2004). Comparison of three priming techniques for onion seed lots differing in initial seed quality. *Seed Science and Technology*, 32, 365-375.

- 9. Cakmak, I. & Horst, W. (1991). Effect of aluminium on lipid peroxidation, superoxide dismutase, catalase and peroxidase activities in root tip of soybean (*Glycine max*). *Plant Physiology*, 83, 463-468.
- Cavalcanti, F. R., Oliveira, J. T. A., Martins-Miranda, A. S., Viégas, R. A. & Silveira, J. A. G. (2004). Superoxide dismutase, catalase and peroxidase activities do not confer protection against oxidative damage in salt-stressed cowpeas leaves. *New Phytologist*, 163, 563-571.
- 11. Chiu, K. Y., Wang, C. S. & Sung, J. M. (1995). Lipid peroxidation and peroxidescavenging enzymes associated with accelerated ageing and hydration of watermelon seeds differing in ploidy. *Plant Physiology*, 94, 441-446.
- 12. Delouche, J. C. & Baskin, C. C. (1973). Accelerated ageing technique for predicting relative storability of seed lots. *Seed Science and Technology*, 1, 427-452.
- 13. El-Araby, M. M. & Hegazi, A. Z. (2004). Responses of tomato seeds to hydro-and osmopriming, and possible relations of some antioxidant enzymes and endogenous polyamine fractions. *Egyptian Journal of Biology*, 6, 81-93.
- 14. Ellis, R. A. & Roberts, E. H. (1981). The quantification of ageing and survival in orthodox seeds. *Seed Science and Technology*, 9, 373-409.
- 15. Fujikura, Y., Kraak, H. L., Basra, A. S. & Karssen, C. M. (1993). Hydropriming, a simple and inexpensive priming method. *Seed Science and Technology*, 21, 639-642.
- 16. Ghasemi-Golezani, K., Aliloo, A. A., Valizadeh, M. & Moghaddam, M. (2008). Effects of different priming techniques on seed invigoration and seedling establishment of lentil (*Lens culinaris* Medik.). *Journal of Food Agriculture and and Environment*, 6, 222-226.
- Ghasemi-Golezani, K., Chadordooz-Jeddi, A., Nasrollahzade, S. & Moghaddam, M. (2010). Effects of hydro-priming duration on seedling vigour and grain yield of pinto bean (*Phaseolus vulgaris* L.) cultivars. *Notulae Botanicae Horti Agrobotanici*, 38(1), 109-113.
- Ghasemi-Golezani, K., Japparpour-Bonyadi, Z., Shafagh-Kolvanagh, J. & Nikpour-Rashidabad, N. (2013). Effects of water stress and hydro-priming duration on field performance of lentil. *International Journal of Farming and Allied Science*, 2, 922-925.
- 19. Giannopolitis, C. & Ries, S. (1977). Superoxid desmutase. I: Occurence in higher plant. *Plant Physiology*, 59, 309-314.
- 20. Hampton, J. G. & TeKrony, D. M. (1995). *Handbook of Vigour Test Methods*. The international Seed Testing Association, Zurikh.
- 21. Hsu, C. C., Chen, C. L., Chen, J. J. & Sung, J. M. (2003). Accelerated ageingenhanced lipid peroxidation in bitter gourd seeds and effects of priming and hot water soaking treatments. *Horticultural Science*, 98, 201-212.
- 22. Irigoyen, J. J., Emerich, D. W. & Sanchez-Diaz, M. (1992). Water stress induced changes in concentrations of proline and total soluble sugars in nodulated alfalfa (*Medicago sativa*) plants. *Physiologia Plantarum*, 84, 55-60.
- 23. Jisha, K. C., Vijayakumari, K. & Puthur, J. T. (2013). Seed priming for abiotic stress tolerance: an overview. *Acta Physiologiae Plantarum*, 35, 1381-1396.
- Jyotsna, V. & Srivastava, A.K. (1998). Physiological basis of salt stress resistance in pigeonpea (*Cajanus cajan* L.)-II. Pre-sowing seed soaking treatment in regulating early seedling metabolism during seed germination. *Plant Physiology and Biochemistry*, 25, 89-94.
- 25. Kamithi, K. D., Wachira, F. & Kibe, A. M. (2016). Effects of different priming methods and priming durations on enzyme activities in germinating chickpea (*Cicer arietinum* L.). *American Journal of Natural and Applied Sciences*, 1(1), 1-9.

- 26. Kaya, M. D., Gamze, O., Atal, M., Yakup, C. & Ozer, K. (2006). Seed treatments to overcome salt and drought stress during germination in sunflower (*Helianthus annuus* L.). *Europian Journal of Agronomy*, 24, 291-295.
- 27. Kibinza, S., Vinel, D., Côme, D., Bailly, C. & Corbineau, F. (2006). Sunflower seed deterioration as related to moisture content during ageing, energy metabolism and active oxygen species scavenging. *Physiologica Plantarum*, 128, 496-506.
- Kibinza, S., Bazin, J., Bailly, C., Farrant, J. M., Corbineau, F. & El-Marrouf Bouteau, H. (2011). Catalase is a key enzyme in seed recovery from ageing during priming. *Plant Science*, 181, 309-315.
- 29. Langens-Grrits, M. M., Miller, W. B., Croes, A. F. & De Klerk, G. J. (2003). Effects of low temperature on dormancy breaking and growth after planting in lily bulblets regenerated in vitro. *Journal of Plant Growth Regulator*, 40, 267-275.
- 30. López, L. V. P., Rodríguez, A. R., Coronado, M. E. S., Hernández, P. E. M., Segovia, A. O. (2016). Effects of hydropriming treatments on the invigoration of aged *Dodonaea viscosa* seeds and water-holding polymer on the improvement of seedling growth in a lava field. *Restoration Ecology*, 24(1), 61-70.
- Matsushima, K. & Sakagami, J. (2012). Effects of seed hydropriming on germination and seedling vigor during emergence of rice under different soil moisture conditions. *American Journal of Plant Science*, 4, 1584-1593.
- 32. Nakano, Y. & Asada, K. (1981). Hydrogen peroxide scavenged by ascrobate-specific peroxidase in spinach chloroplast. *Plant Cell Physiology*, 22, 867-880.
- 33. Nascimento, W.M. (2012). Muskmelon seed germination and seedling development in response to seed priming. *Scientia Agricola*, 60(1), 71-75.
- 34. Rahnama-Ghahfarokhi, A. & Tavakkol-Afshari, R. (2007). Methods for dormancy breaking and germination of galbanum seeds (*Ferula gummosa*). Asian Journal of *Plant Science*, 6, 611-616.
- 35. Sacala, E. & Demczuk, A. (2016). Effect of laser and hydropriming of seeds on some physiological parameters in sugar beet. *Journal of Elementology*, 21(2), 527-538.
- 36. Sallam, H. A. (1999). Effect of some seed-soaking treatments on growth and chemical components on faba bean plants under saline conditions. *Annals of Agricultural Sciences*, 44, 159-171.
- 37. Sepehri, A. & Rouhi, H. R. (2016). Enhancement of seed vigor performance in aged groundnut (*Arachis hypogaea* L.) seeds by sodium nitroprusside under drought stress. *The Philippine Agricultural Scientist*, 99 (4), 339-347.
- Sharma, A. D., Rathore, S. V. S., Srinivasan, K. & Tyagi, R. K. (2014). Comparison of various seed priming methods for seed germination, seedling vigour and fruit yield in okra (*Abelmoschus esculentus* L. Moench). *Scientia Horticulture*, 165, 75-81.
- 39. Sung, J. M. & Chiu, K. Y. (1995). Hydration effects on seedling emergence strength of watermelon seed differing in ploidy. *Plant Science*, 110, 21-26.
- 40. Wattanakulpakin, P., Photchanachai, S., Ratanakhanokchai, K., Kyu, K. L., Ritthichai, P. & Miyagawa, S. (2012). Hydropriming effects on carbohydrate metabolism, antioxidant enzyme activity and seed vigor of maize (*Zea mays L.*). *African Journal of Biotechnology*, 11, 3537-3547.
- 41. Wilson, D. O. & McDonald, M. B. (1986). The lipid peroxidation model of seed deterioration. *Seed Science and Technology*, 14, 269-300.
- 42. Yan, M. (2015). Hydropriming promotes germination of aged napa cabbage seeds. *Seed Science and Technology*, 43(2), 303-307.
- 43. Yan, M. (2016). Hydro-priming increases seed germination and early seedling growth in two cultivars of Napa cabbage (*Brassica rapa* subsp. pekinensis) grown under salt stress. *The Journal of Horticultural Science & Biotechnology*, 91(4), 1-6.